

Very Fast Kinetics

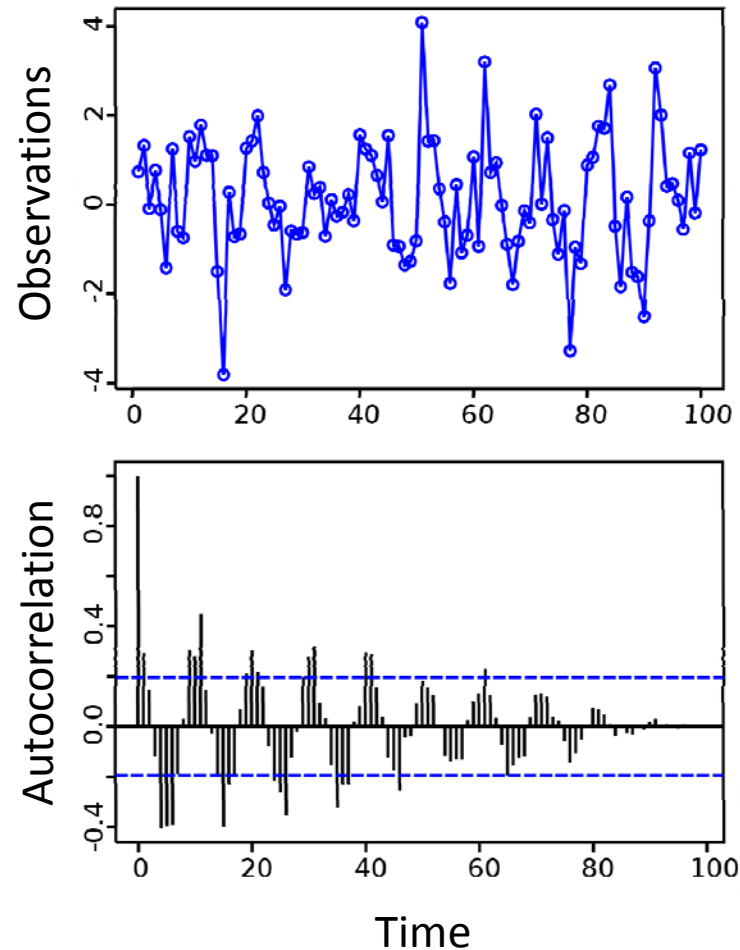
- What is “very fast?”
 - Stopped flow methods: $1+ \text{ms}^{-1}$ rates
 - What about $10\text{-}100 \mu\text{s}^{-1}$ rates?
- Examples:
 - Very fast protein folding
 - pH neutralization
 - Some ion binding (e.g. Ca^{2+})

Fluctuation Methods

- **Idea:** Measure some quantity as it changes over time
 - System is at equilibrium, but changes still occur
 - Fluctuations will reflect kinetics
- **Challenges:**
 - Need sensitive equipment to measure small changes
 - Need a system that is theoretically (and practically) tractable

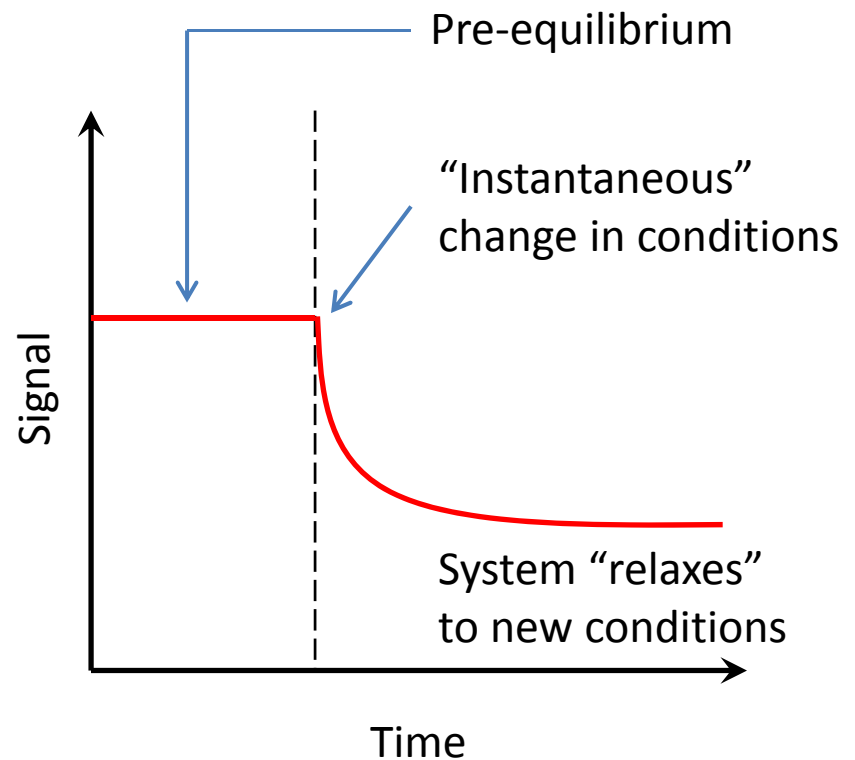
Fluctuation Methods: Examples

- Methods
 - Refractive index
 - Concentration
 - Light scattering
 - Pressure
- **Autocorrelation:** how an observed quantity correlates to itself (typically over time)



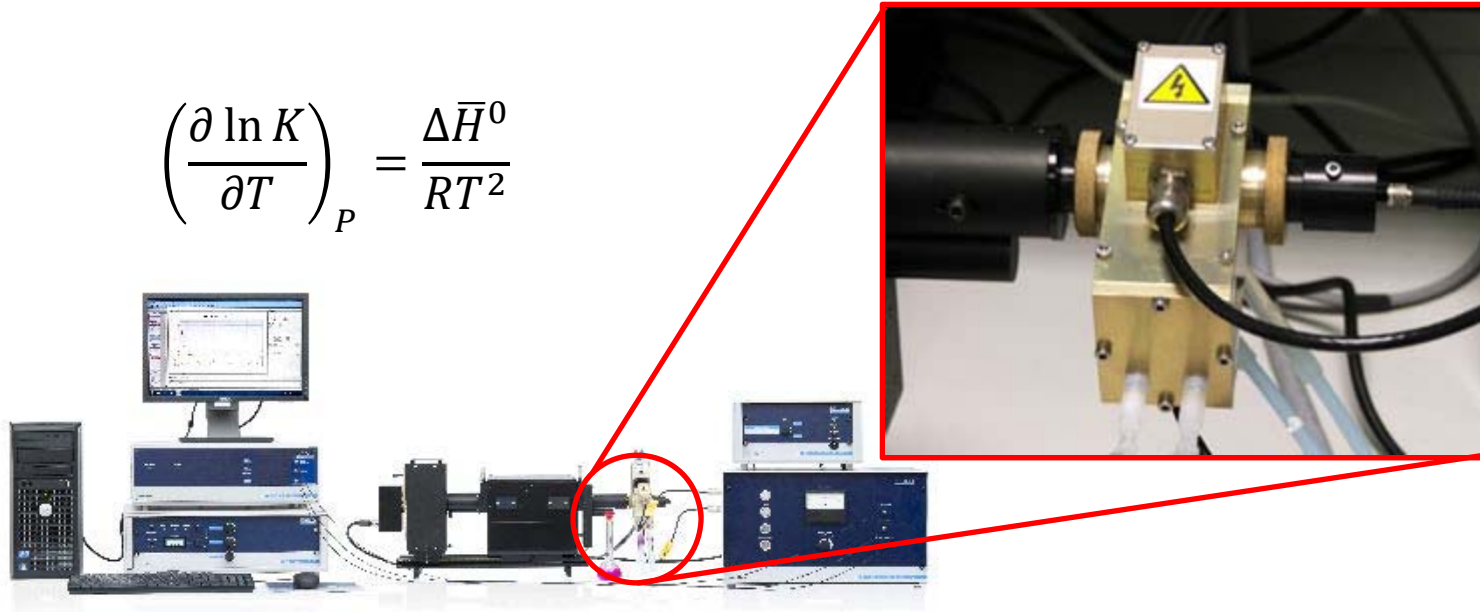
Relaxation/Perturbation Methods

- **Idea:** Start at equilibrium, then change conditions (no new intermediates)
- New equilibrium will be established → observe change over time
- **Critical:** Kinetics will reflect new conditions, not old conditions



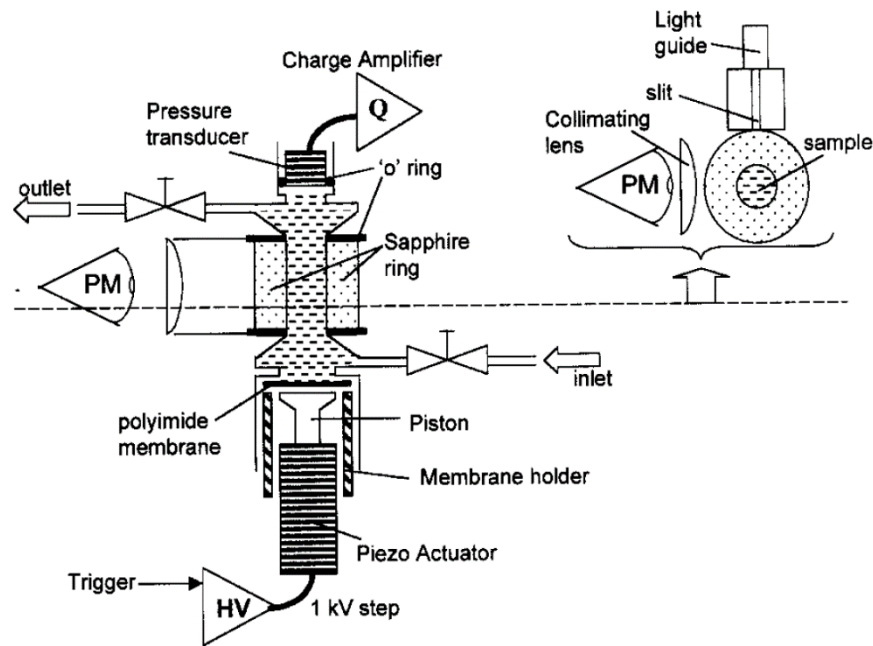
Method 1: Temperature Jump

$$\left(\frac{\partial \ln K}{\partial T}\right)_P = \frac{\Delta \bar{H}^0}{RT^2}$$



- Electrical discharge, laser pulse rapidly increases temperature (small sample volume)

Method 2: Pressure Jump

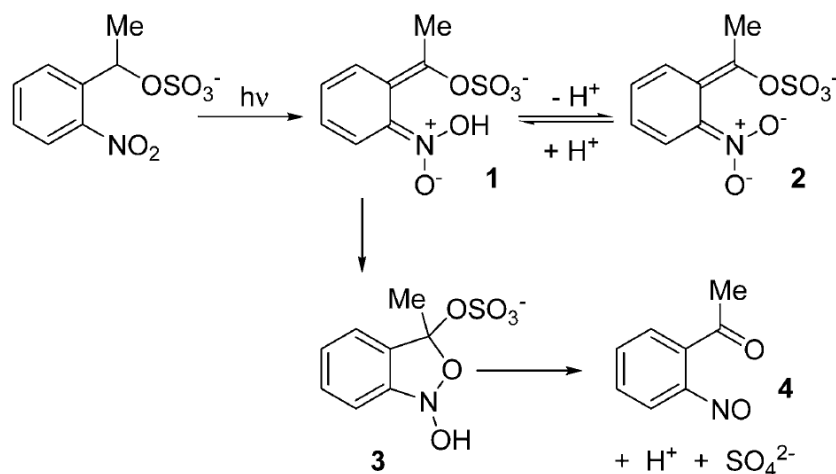


$$\left(\frac{\partial \ln K}{\partial P}\right)_T = -\frac{\Delta \bar{V}^0}{RT}$$

- Diaphragm bursts to increase (or decrease) pressure of system
- To keep change in K small, $\Delta P \approx 10^2 - 10^3$ atm

Method 3: Flash Photolysis

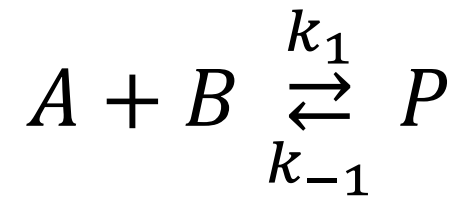
Scheme 1. Proposed Reaction Mechanism for Photolysis of 1-(2-Nitrophenyl)ethyl Sulfate¹⁴



- Rapid flash of light (or laser pulse) releases H^+ to change pH
- Other “caged” molecules exist (e.g. ATP)


Relaxation Kinetics Example

- Binding reaction:



Why Can We Ignore x^2 ?

(or ΔP^2)

- Consider equilibrium:
 - Let $A_{eq} = 10 \text{ mM}$ and $B_{eq} = 50 \text{ mM}$
 - If small perturbation, x is 1 mM
- Then:
 - $A_{eq}B_{eq} = 500 \text{ mM}^2$
 - $(A_{eq} + B_{eq})x = 60 \text{ mM}^2$
 - $x^2 = 1 \text{ mM}^2$  This is much smaller than the other two

Implications of Relaxation Kinetics

- Remember kinetic rates (and equilibrium) concentrations are from new conditions!!
- Curves will always be exponential, because x^2 will always be small if perturbation is small
- Determining τ at multiple conditions (e.g. A_{eq} , B_{eq} and P_{eq}) can reveal rate constants

Relaxation Time Constants

TABLE 7.4 Relaxation Times for Reactions Involving Single Steps

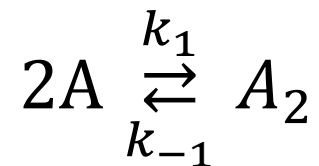
Mechanism	Relaxation time*
$A \xrightleftharpoons[k_{-1}]{k_1} B$	$\tau = \frac{1}{k_1 + k_{-1}}$
$A + B \xrightleftharpoons[k_{-1}]{k_1} P$	$\tau = \frac{1}{k_{-1} + k_1([\bar{A}] + [\bar{B}])}$
$A + B + C \xrightleftharpoons[k_{-1}]{k_1} P$	$\tau = \frac{1}{k_{-1} + k_1([\bar{A}][\bar{B}] + [\bar{B}][\bar{C}] + [\bar{A}][\bar{C}])}$
$A + B \xrightleftharpoons[k_{-1}]{k_1} P + Q$	$\tau = \frac{1}{k_1([\bar{A}] + [\bar{B}]) + k_{-1}([\bar{P}] + [\bar{Q}])}$
$2A \xrightleftharpoons[k_{-1}]{k_1} A_2$	$\tau = \frac{1}{4k_1[\bar{A}] + k_{-1}}$

* $[\bar{A}]$, $[\bar{B}]$, etc., represent the equilibrium concentrations after the perturbation.

Source: M. Eigen and L. De Maeyer, in *Investigation of Rates and Mechanisms of Reactions*, 3d ed., vol. 6, part 2, ed. G. G. Hammes (New York: Wiley-Interscience, 1974), chapter 3.

Example: Dimerization Kinetics in DNA

- Dimerization scheme:



- Time constant:

$$\tau = \frac{1}{4k_1A_{eq} + k_{-1}}$$

- Modified time constant:

$$\frac{1}{\tau^2} = k_{-1}^2 + 8k_1k_{-1}A_{total}$$

Strategy: Relaxation Kinetics

- For individual points (to determine τ):
 - Always first-order kinetics (exponential)
 - Plot $\ln[A]$ vs. $t \rightarrow$ slope will be $\pm \frac{1}{\tau}$
- Determine how τ varies with A_{eq} , B_{eq} , etc.
 - Figure out how to linearize the function to relate slope, intercept to kinetic parameters
- **Remember:** Linearization is bad for real-life situations; use nonlinear least squares